Molecular Markers of Eosinophilopoesis at Birth: Kinetics of Cord Blood GATA-1, MBP and IL-5 Receptor Expression

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Abstract

Objective/Purpose: Using colony assays and flow cytometry, we have recently shown that eosinophil/basophil (Eo/B) progenitor phenotype and function are associated with atopic risk at birth and infant clinical outcomes. The current study aimed to utilize real-time polymerase chain reaction (Q-PCR) to ascertain the kinetic patterns of expression of CB Eo/B-lineage specific genes, GATA-1, MBP and IL-5Rα in order to develop molecular markers of Eo/B differentiation.

Methods: CB non-adherent mononuclear cells (NANMCs) were isolated from random fresh and frozen samples, and incubated in the presence of IL-5 (1 ng/mL), IL-4, and IL-3 (72 and 48 hr post-stimulation). RNA was isolated, reverse transcribed, and expression of IL-5Rα, GATA-1, and MBP were determined utilizing comparative Q-PCR in a multiplex reaction. The relative expression ratios between stimulated and un-stimulated cells were calculated using the delta-delta Ct method.

Findings: Stimulation with IL-5 resulted in an up-regulation of GATA-1 expression, which peaked between 24 and 48hrs. In contrast, MBP was up-regulated in a slowly progressive pattern, with maximal up-regulation at 72hr, while there was a stable, minor down-regulation of IL-5Rα. In keeping with these molecular kinetics, Eo/B colony-forming cells, grown in 14 day methycellulose cultures, were found to be present in relation to timing of GATA-1 expression.

Relevance: Multiplex Q-PCR analysis of mRNA from CB mononuclear cells stimulated with IL-5 demonstrates sequential expression of critical lineage-specific events, can be used as a surrogate, molecular marker of CB Eo/B differentiation in basic research and clinical studies.

Introduction

• We have previously identified that the number and phenotype of cord blood (CB) eosinophil/basophil (Eo/B) progenitor cell relate to atopic risk and infant clinical outcomes
• All previous evaluations utilized either 14 day methycellulose colony assays or flow cytometry analysis
• We have previously demonstrated the kinetics of GATA-1 expression in CB mononuclear cells (CBMNC) with IL-5 stimulation, and shown these to be in concert with subsequent Eo/B colony forming units (CFUs) in methycellulose culture.
• We have not examined the expression of other molecular markers of Eo/B lineage commitment such as the IL-5R or Major Basic Protein (MBP).

Objective

• To ascertain the kinetic patterns of expression of CB Eo/B-lineage specific genes: GATA-1, MBP and IL-5Rα via a multiplex Q-PCR reaction, in order to develop molecular markers of Eo/B differentiation.

Results

• Cord blood (CB) non-adherent mononuclear cells (NANMCs) were isolated from random fresh and frozen samples, and incubated in the presence of rhIL-5 (1 ng/mL) (10 million cells included for each condition)
• At 24, 48 and 72 hr post-stimulation:
  • RNA extracted using RiNAeasy® Mini-Kit columns (Qiagen) according to manufacturer’s instructions
  • DNA contamination removed using DNA Free® kit® containing DNase1 buffer and DNase1 mix (Ambion) according to manufacturer’s instructions
  • Total RNA in each sample was quantified using UV spectrophotometer
  • 2 mg RNA reverse-transcribed for each sample (volume calculated according to Total RNA quantification)
  • Reverse-transcription completed with 2.97 ml random hexamer primers and 0.03 ml oligo (dT) primer (both 100ng/ml)
• Expression of IL-5Rα, GATA-1, and MBP were determined utilizing comparative Q-PCR in a multiplex reaction (Stratagene MX4000); Housekeeping gene was GAPDH
• Normalized relative expression ratios between stimulated and un-stimulated cells were calculated using the delta-delta Ct method (2-∆∆Ct)

Discussion

The patterns of expression of each of the genes evaluated were consistent with our expectations, since GATA-1 is known to be an early, essential transcription factor in eosinophil lineage commitment. In addition, our findings in the current experiment were in concordance with previous evaluations in this laboratory of the kinetics of GATA-1 expression over time in CBMNC.

MBP is produced much later in eosinophilopoiesis, and is a marker of mature Eo commitment. That the up-regulation seen of MBP mRNA was slow and steady with a 4 fold increase by 72 hrs was in keeping with previous evaluations demonstrating MBP production (i.e. product) peaking after 12 days of culture. In addition, MBP expression was positively correlated with later Eo/B CFU development in 2 wk culture.

It has been previously shown that IL-5 has the ability to downregulate its own receptor; we have shown that while total IL-5Rα expression patterns remain stable, there is differential expression of the transmembrane and soluble isoforms: progressive up-regulation of the soluble isofrom is accompanied by downregulation of the transmembrane.

Conclusions

• mRNA of GATA-1, IL-5Rα, and MBP can be evaluated in a Multiplex Q-PCR reaction.
• Analysis of mRNA from cord blood nonmononuclear cells stimulated with rhIL-5 demonstrates sequential expression of critical Eo/B lineage-specific events
• Q-PCR analysis of the expression of these genes can be used as a surrogate, molecular marker of CB Eo/B differentiation in basic research and clinical studies

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